

PLANT GENETICS

Early allopolyploid evolution in the post-Neolithic *Brassica napus* oilseed genome

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Oilseed rape (*Brassica napus* L.) was formed ~7500 years ago by hybridization between *B. rapa* and *B. oleracea*, followed by chromosome doubling, a process known as allopolyploidy. Together with more ancient polyploidizations, this conferred an aggregate 72× genome multiplication since the origin of angiosperms and high gene content. We examined the *B. napus* genome and the consequences of its recent duplication. The constituent A_n and C_n subgenomes are engaged in subtle structural, functional, and epigenetic cross-talk, with abundant homeologous exchanges. Incipient gene loss and expression divergence have begun. Selection in *B. napus* oilseed types has accelerated the loss of glucosinolate genes, while preserving expansion of oil biosynthesis genes. These processes provide insights into allopolyploid evolution and its relationship with crop domestication and improvement.

The Brassicaceae are a large eudicot family (1) and include the model plant *Arabidopsis thaliana*. Brassicas have a propensity for genome duplications (Fig. 1) and genome mergers (2). They are major contributors to the human diet and were among the earliest cultigens (3).

B. napus (genome A_nA_nC_nC_n) was formed by recent allopolyploidy between ancestors of *B. oleracea* (Mediterranean cabbage, genome C₀C₀) and *B. rapa* (Asian cabbage or turnip, genome A_rA_r) and is polyphyletic (2, 4), with spontaneous formation regarded by Darwin as an example of unconscious selection (5). Cultivation began in Europe during the Middle Ages and spread worldwide. Diversifying selection gave rise to oilseed rape (canola), rutabaga, fodder rape, and kale morphotypes grown for oil, fodder, and food (4, 6).

The homozygous *B. napus* genome of European winter oilseed cultivar ‘Darmor-*bzh*’ was assembled with long-read [>700 base pairs (bp)] 454 GS-FLX+ Titanium (Roche, Basel, Switzerland) and Sanger sequence (tables S1 to S5 and figs. S1 to S3) (7). Correction and gap filling used 79 Gb of Illumina (San Diego, CA) HiSeq sequence. A final assembly of 849.7 Mb was obtained with SOAP (8) and Newbler (Roche), with 89% nongapped se-

quence (tables S2 and S3). Unique mapping of ~5× nonassembled 454 sequences from *B. rapa* (‘Chiifu’) or *B. oleracea* (‘TO1000’) assigned most of the 20,702 *B. napus* scaffolds to either the A_n (8294) or the C_n (9984) subgenomes (tables S4 and S5 and fig. S3). The assembly covers ~79% of the 1130-Mb genome and includes 95.6% of *Brassica* expressed sequence tags (ESTs) (7). A single-nucleotide polymorphism (SNP) map (tables S6 to S9 and figs. S4 to S8) genetically anchored 712.3 Mb (84%) of the genome assembly, yielding pseudomolecules for the 19 chromosomes (table S10).

The assembled C_n subgenome (525.8 Mb) is larger than the A_n subgenome (314.2 Mb), consistent with the relative sizes of the assembled C₀ genome of *B. oleracea* (540 Mb, 85% of the ~630-Mb genome) and the A_r genome of *B. rapa* (312 Mb, 59% of the ~530-Mb genome) (9–11). The *B. napus* assembly contains 34.8% transposable elements (TEs), less than the 40% estimated from raw reads (tables S11 to S14) (7), with asymmetric distribution in the A_n and C_n subgenomes (table S12) as in the progenitor genomes (9–11). A small TE fraction has proliferated since *B. napus* separated from its progenitors (7), at lower rates in the *B. napus* subgenomes than the corresponding progenitor genomes (table S14 and figs. S9 and S10).

The *B. napus* genome contains 101,040 gene models estimated from 35.5 Gb of RNA sequencing (RNA-seq) data (tables S15 and S16) in combination with ab initio gene prediction, protein and EST alignments, and transposon masking (7). Of these, 91,167 were confirmed by matches with *B. rapa* and/or *B. oleracea* predicted proteomes. Genes are abundant in distal euchromatin but sparse near centromeric heterochromatin (Fig. 2). RNA-seq data revealed alternative splicing in 48% of genes, with frequent intron retention (62%) and rare exon skipping (3%) (tables S17 and S18 and fig. S11).

The *B. napus* A_n and C_n subgenomes are largely colinear to the corresponding diploid A_r and C₀

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